

## New 3(2H)-pyridazinone derivatives: synthesis and affinity towards $\alpha_1$ AR subtypes and 5HT<sub>1A</sub> receptors

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**Summary** — The synthesis and the evaluation of the radioreceptor binding affinity of a series of 3(2H)-pyridazinone derivatives is reported. Their affinity towards the  $\alpha_1$  receptor, three  $\alpha_1$ -AR subtype ( $\alpha_{1A}$ / $\alpha_{1B}$ / $\alpha_{1D}$ ) receptors and the 5HT<sub>1A</sub> receptor has been determined. The results of the affinity on  $\alpha_1$ -AR subtypes and the 5HT<sub>1A</sub>/ $\alpha_1$  selectivity are discussed.

**structure–activity relationship /  $\alpha_1$  and 5HT<sub>1A</sub> receptor / pyridazinone derivative / arylpiperazine**

### Introduction

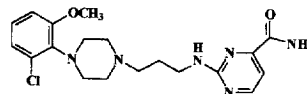
Pharmacological evidence and recent molecular cloning studies have demonstrated that the  $\alpha_1$ -AR is not a homogeneous population and three  $\alpha_1$ -AR subtypes have been characterized by functional, radioligand binding and molecular biology techniques. These are usually referred to as  $\alpha_{1A}$  ( $\alpha_{1A}$ ),  $\alpha_{1B}$  ( $\alpha_{1B}$ ), and  $\alpha_{1D}$  ( $\alpha_{1D}$ ) adrenoceptors where the lower case subscripts are used for recombinant receptors and the upper case subscripts for receptors in native tissues [1].

Recently, a great deal of attention has been paid towards obtaining  $\alpha_1$ -antagonists with high selectivity towards only one type of subreceptor. This interest arises from the observation that only compounds with selectivity show activity on the lower tract of the urethra [2].

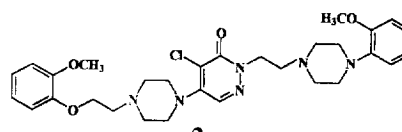
Therefore with the hope of obtaining compounds which are highly selective towards only one  $\alpha_1$  subreceptor, we synthesized compound **3** in which the 3(2H)-pyridazinone ring was linked in the 2-position with the 1-aryl piperazine through a chain of three carbon atoms, and in the 6-position with a phenyl group. The 3(2H)-pyridazinone ring was chosen since its derivatives show high adrenergic activity, and since compounds like **1** [2] with a 1-phenyl-4-piperazin alkyl fragment show a high selectivity towards the  $\alpha_{1A}$  subtype. Furthermore, we synthesized com-

pound **7** in which the phenyl group was substituted with a furoylpiperazinyl group and compounds **4–6** in which the 3(2H)-pyridazinone ring was linked in 2-position with the differently substituted 1-aryl piperazine through a chain of two carbon atoms, since we were interested in determining the influence of these groups, present in many  $\alpha_1$  antagonists, towards the  $\alpha_1$  subreceptors.

It is well known that the  $\alpha_1$  adrenoceptor ( $\alpha_1$ -AR) is a member of the super family of G-protein-coupled receptors as well as the 5HT<sub>1A</sub> receptor. In spite of their completely distinct pharmacology, these receptors show common features in their binding sites [3, 4]. Therefore it was interesting to determine the affinity of all the compounds towards the 5HT<sub>1A</sub> receptor and their selectivity (5HT<sub>1A</sub>/ $\alpha_1$ ). Finally, the pharmacological results obtained for compounds **3–7** were compared with the affinity towards  $\alpha_1$ -AR subtypes and towards the 5HT<sub>1A</sub> receptor found for compound **2**, which was synthesized previously in our laboratory [5].



**1**



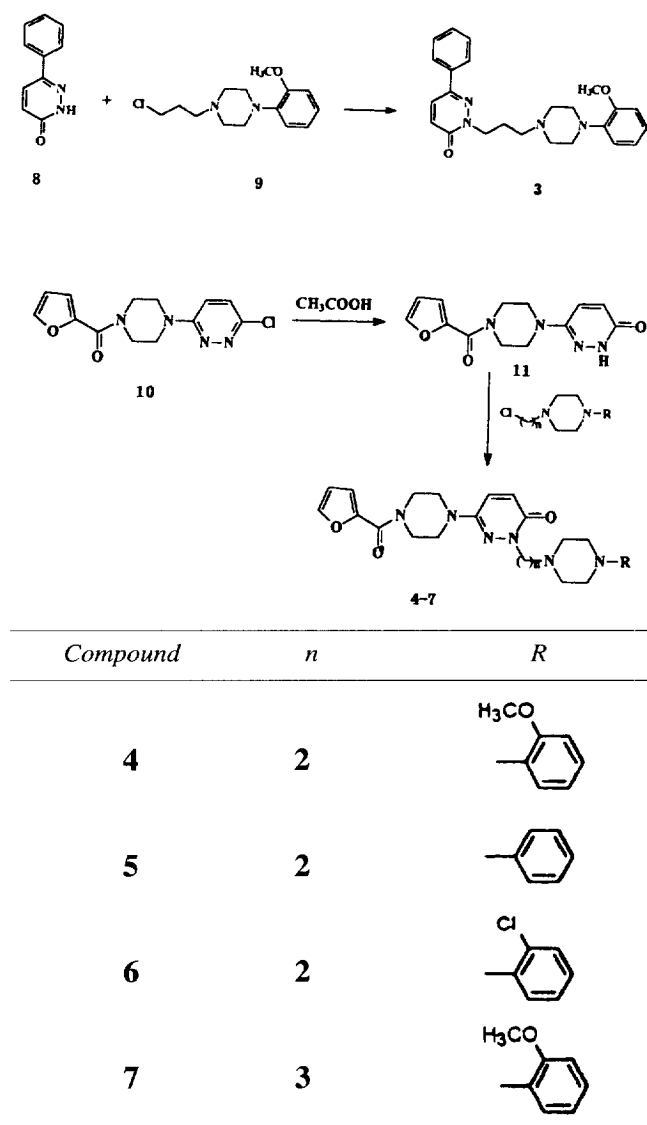
**2**

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Dedicated to Professor Mario Piattelli on the occasion of his 70th birthday.

## Chemistry

Compound **3** was prepared by alkylation of 6-phenyl-3(2*H*) pyridazinone **8** with 4-(2-methoxyphenyl)-1-(3-chloropropyl)piperazine **9** in dry methanol and potassium hydroxide pellets. Compound **11** was prepared by alkylation of 1-(2-furoyl)piperazine with 3,6-dichloropyridazine (compound **10**) followed by hydrolysis with glacial acetic acid. Starting from compound **11**, by alkylation with the appropriate 1-aryl-piperazinyl alkyl halide in dry ethanol and sodium hydroxide pellets in equimolar ratio, the corresponding compounds **4–7** were obtained (scheme 1).



Scheme 1.

## Results and discussion

From the pharmacological results reported in tables I and II, compound **3**, in which a [4-(2-methoxyphenyl)-1-piperazinyl]propyl system is linked in the 2-position and a phenyl ring in the 6-position of the pyridazinone ring, shows a higher affinity towards the  $\alpha_1$  adrenoceptor (table II), particularly towards the  $\alpha_{1a}$  subtype (table I) compared with compound **7** in which the phenyl group was substituted by the 4-(2-furoyl)-1-piperazinyl system. Compound **4**, obtained by shortening the alkyl chain linking the 6-[4-(2-furoyl)-1-piperazinyl]-3(2*H*) pyridazinone to the 4-(2-methoxyphenyl)-1-piperazine moiety of compound **7** by one unit, shows a slight increase in affinity towards the  $\alpha_{1a}$  subtype with an increase in selectivity towards the  $\alpha_{1b}$  subtype. Furthermore, compounds **4** and **7** have a similar affinity towards the  $\alpha_{1b}$  subtype.

In compounds **5** and **6**, in which the methoxy group was eliminated or substituted with a chlorine atom respectively, both affinity and selectivity decreased.

Table I. Affinity towards different  $\alpha_1$  adrenoceptor subtypes.

Compound	$\alpha_{1a}$ $K_i$ (nM)	$\alpha_{1b}$ $K_i$ (nM)	$\alpha_{1d}$ $K_i$ (nM)
3	1.8	47.1	2.1
4	4.7	793.5	11.5
5	94.6	111.6	445.9
6	11.4	41.9	4.9
7	24.5	734.4	18.3
5-Methylurapidil	2.0	775.0	27.4

Table II. Affinity towards  $\alpha_1$  and 5HT<sub>1A</sub> receptor types.

Compound	$\alpha_1$ $K_i$ (nM)	5HT <sub>1A</sub> $K_i$ (nM)	Ratio $K_i$ / 5HT <sub>1A</sub> / $\alpha_1$
3	17.5	60.0	3.42
4	282.8	5825.2	20.60
5	1261.9	6427.6	5.09
6	176.3	731.7	4.15
7	118.0	2292.3	19.42
5-Methylurapidil	28.0	1.2	0.04

In table III, the affinity of pyridazinone derivative **2** towards  $\alpha_1$  subtypes ( $\alpha_{1a}/\alpha_{1b}/\alpha_{1d}$ ) and 5HT<sub>1A</sub> receptors is reported. These pharmacological results can be compared with the pharmacological data obtained for compounds **3–7** to show that compounds **2** and **4** possess a similar affinity towards  $\alpha_{1a}$  and  $\alpha_{1d}$ , while the affinity towards  $\alpha_{1b}$  is higher for compound **2**. Therefore the substitution of the 4-(2-methoxyphenoxyethyl)-1-piperazinyl fragment (compound **2**) with the 1-(2-furoyl)piperazine group (compound **4**) increases the selectivity.

The affinity of all the compounds towards the 5HT<sub>1A</sub> receptor and 5HT<sub>1A</sub>/ $\alpha_1$  selectivity was also determined (table II). These compounds show a low affinity for this receptor, except compound **3**; the highest selectivity (5HT<sub>1A</sub>/ $\alpha_1$ ) was found for compounds **4**.

Compounds **4** in particular show an  $\alpha_1$ -AR receptor binding profile similar to the  $\alpha_{1a}$ -selective antagonist 5-methylurapidil, with the remarkable advantage of a more than 1000-fold selectivity when the  $\alpha_{1a}$  and 5-HT<sub>1A</sub> receptors are considered.

In conclusion, these results provide more information on the affinity and selectivity of new pyridazinone derivatives towards the  $\alpha_1$ -AR subtype and on the 5-HT<sub>1A</sub> receptors.

**Table III.** Affinity towards subtypes  $\alpha_{1a}/\alpha_{1b}/\alpha_{1d}$  and 5HT<sub>1A</sub> receptors for compound **2**.

Compound	$K_i$ (nM) <sup>a</sup>			
	$\alpha_{1a}$	$\alpha_{1b}$	$\alpha_{1d}$	5HT <sub>1A</sub>
<b>2</b>	7.45	130	31	270
Prazosin	0.2	0.5	0.3	–
Trazodone	–	–	–	120

<sup>a</sup>The  $K_i$  binding data were calculated as described by De Blasi [13].

## Experimental protocols

### Biological methods

#### Radioligand binding assay at cloned bovine $\alpha_{1a}$ and hamster $\alpha_{1b}$ adrenoreceptors expressed in COS-7 cells

[<sup>3</sup>H]Prazosin binding to cloned  $\alpha_1$  adrenoreceptor subtypes was performed in COS-7 cells (CV-1 monkey kidney epithelial cells) transiently expressing bovine  $\alpha_{1a}$  and hamster  $\alpha_{1b}$  adrenoreceptor subtypes were carried out by S Cotecchia [6], as previously described [7]. COS-7 cell membranes (35  $\mu$ g proteins) were

incubated in 50 mM Tris–HCl pH 7.4, containing 10  $\mu$ g pargyline and 0.1% ascorbic acid, with 0.1–0.4 nM [<sup>3</sup>H]prazosin, in a final volume of 0.22 mL for 30 min at 25 °C, in the absence or the presence of competing drugs. Non-specific binding was determined in the presence of 100  $\mu$ M phentolamine.

The incubation was stopped by addition of ice-cold Tris–HCl buffer and rapid filtration through 0.2% polyethyleneimine pretreated Whatman GF/B or Schleicher and Schuell GF 52 filters.

#### Radioligand binding assay at cloned $\alpha_{1d}$ adrenoreceptors expressed in CHO cells

Binding to cloned human  $\alpha_{1d}$  adrenoreceptors was performed in membranes from CHO cells (Chinese hamster ovary cells) transfected by electroporation with DNA expressing the gene encoding the  $\alpha_{1d}$  adrenoreceptor subtype. Cloning and stable expression of the human  $\alpha_{1d}$  adrenoreceptor gene was performed as previously described [8].

CHO cell membranes (30  $\mu$ g proteins) were incubated in 50 mM Tris–HCl, pH 7.4, with 0.1–0.4 nM [<sup>3</sup>H]prazosin, in a final volume of 1.02 mL for 30 min at 25 °C, in the absence or the presence of competing drugs. Non-specific binding was determined in the presence of 10  $\mu$ M phentolamine.

The incubation was stopped by addition of ice-cold Tris–HCl buffer and rapid filtration through 0.2% polyethyleneimine pretreated Whatman GF/B or Schleicher and Schuell GF 52 filters.

#### Radioligand binding assay on native receptors

Radioligand binding studies on  $\alpha_1$  adrenergic and 5-HT<sub>1A</sub> serotonergic receptors were performed as previously described [9, 10] with minor modifications.

**Membrane preparation.** Male rats were killed by cervical dislocation and cerebral cortex ( $\alpha_1$  adrenergic receptor) and hippocampus (5-HT<sub>1A</sub> serotonergic receptor) immediately frozen and stored at –70 °C until use. Tissues were homogenized (2 x 20 sec) in 50 vol of cold Tris–HCl, pH 7.4, using a Politron homogenizer (speed 7). Homogenates were centrifuged at 49 000 g for 10 min, resuspended in 50 vol of the same buffer, incubated at 37 °C for 15 min and centrifuged and resuspended twice more. The final pellets were resuspended in 100 vol of Tris–HCl buffer pH 7.4, containing 10  $\mu$ M pargyline and 0.1% ascorbic acid.

**Binding assay.** Membranes were incubated with 0.2–0.4 nM [<sup>3</sup>H]prazosin ( $\alpha_1$  adrenergic receptor) or 1 nM [<sup>3</sup>H]8-OH-DPAT (5-HT<sub>1A</sub>) in a final vol of 1 mL for 30 min at 25 °C, in the absence or the presence of competing drugs. Non-specific binding was determined in the presence of 10  $\mu$ M phentolamine ( $\alpha_1$ ) or 10  $\mu$ M 5-HT (5-HT<sub>1A</sub>). The incubation was stopped by addition of ice-cold Tris–HCl buffer and rapid filtration through 0.2% polyethyleneimine pretreated Whatman GF/B or Schleicher and Schuell GF 52 filters.

### Chemistry

Melting points were determined using a Kofler hot-stage apparatus and are uncorrected. The NMR spectra were recorded with a Bruker AC 200 MHz instrument in the solvent indicated below. The chemical shift values (ppm) are relative to tetramethylsilane as the internal standard. Elemental analyses are within  $\pm 0.4\%$  of the theoretical values. Precoated Kieselgel 60 F<sub>254</sub> plates (Merck) were used for TLC.

**2-[[4-(2-Methoxyphenyl)-1-piperazinyl]propyl]-6-phenyl-3(2H)-pyridazinone 3**

To 50 mL of dry methanol was added 0.17 g ( $3.1 \times 10^{-3}$  mol) of potassium hydroxide pellets and 0.5 g ( $2.9 \times 10^{-3}$  mol) of 6-phenyl-3(2H)-pyridazinone **8** [11]. The mixture was refluxed for 15–20 min, and then 0.76 g ( $2.9 \times 10^{-3}$  mol) of 4-(2-methoxyphenyl)-1-(3-chloropropyl)piperazine **9** [12] was added, and was refluxed under stirring for 8 h. The mixture was concentrated under reduced pressure and the residue was purified by flash chromatography using  $\text{CH}_2\text{Cl}_2/\text{EtOH}$  (96:4) as eluent, giving a dense oil. Yield: 40%.  $^1\text{H-NMR}$  ( $\text{CDCl}_3$ )  $\delta$ : 2.1–2.3 (2H, m,  $\text{CH}_2$ ), 2.55 (2H, t,  $J = 6$  Hz,  $\text{CH}_2$ ), 2.65–2.75 (4H, m, H-piperazinic), 3.05–3.15 (4H, m, H-piperazinic), 3.85 (3H, s,  $\text{OCH}_3$ ), 4.4 (2H, t,  $J = 6$  Hz,  $\text{CH}_2$ ), 6.8–7.05 (5H, m, 4H-aromatic, 1H-pyridazinonic), 7.4–7.5 (3H, m, H-aromatic), 7.65 (1H, d,  $J = 9.5$  Hz, H-pyridazinonic), 7.75–7.85 (2H, m, H-aromatic). The corresponding hydrochloride had mp = 166–170 °C.

**General method for compounds 4–7**

**2-[[4-(2-Methoxyphenyl)-1-piperazinyl]ethyl]-6-[4-(2-furoyl)-1-piperazinyl]-3(2H)-pyridazinone 4.** To 20 mL of dry ethanol was added 0.16 g ( $4.0 \times 10^{-3}$  mol) of sodium hydroxide pellets and 1.1 g ( $4.0 \times 10^{-3}$  mol) of 6-[4-(2-furoyl)-1-piperazinyl]-3(2H)-pyridazinone **11**; this mixture was refluxed for 30 min. Then 1.0 g ( $4.0 \times 10^{-3}$  mol) of the 4-(2-methoxyphenyl)-1-(2-chloroethyl)piperazine [12] dissolved in dry ethanol was added, and this mixture was refluxed under stirring for 5 h. After evaporation under reduced pressure, the residue was purified by chromatography silica gel using as eluent a stepwise gradient of ethanol (0–3%) in  $\text{CH}_2\text{Cl}_2$ . Yield: 40%.  $^1\text{H-NMR}$  ( $\text{CDCl}_3$ )  $\delta$ : 2.7–2.9 (6H, m, 4H-piperazinic,  $\text{CH}_2$ ), 3.0–3.15 (4H, m, H-piperazinic), 3.3–3.4 (4H, m, H-piperazinic), 3.9 (3H, s,  $\text{OCH}_3$ ), 3.95–4.1 (4H, m, H-piperazinic), 4.3 (2H, t,  $J = 6$  Hz,  $\text{CH}_2$ ), 6.5 (1H, m, H-furanic), 6.8–7.1 (7H, m, 4H-aromatic, 2H-pyridazinonic, 1H-furanic), 7.5 (1H, s, H-furanic). The corresponding hydrochloride had mp = 65–68 °C.

**2-[[4-(Phenyl)-1-piperazinyl]ethyl]-6-[4-(2-furoyl)-1-piperazinyl]-3(2H)-pyridazinone 5.** This compound was prepared by alkylation of **11** with 4-(phenyl)-1-(2-chloroethyl)piperazine [12], purified by chromatography silica gel using  $\text{CH}_2\text{Cl}_2/\text{EtOH}$  (95:5) as eluent, and a dense oil was obtained. Yield: 40%.  $^1\text{H-NMR}$  ( $\text{CDCl}_3$ )  $\delta$ : 2.65–2.75 (4H, m, 4H-piperazinic), 2.85 (2H, t,  $J = 6$  Hz,  $\text{CH}_2$ ), 3.15–3.25 (4H, m, H-piperazinic), 3.3–3.4 (4H, m, H-piperazinic), 3.8–3.9 (4H, m, H-piperazinic), 4.2 (2H, t,  $J = 6$  Hz,  $\text{CH}_2$ ), 6.5 (1H, m, H-furanic), 6.8–6.95 (5H, m, 5H-aromatic), 7.0–7.1 (2H, m, H-aromatic), 7.2–7.3 (1, m, H-aromatic), 7.5 (1H, s, H-furanic). The corresponding hydrochloride had mp = 115–120 °C.

**2-[[4-(2-Chlorophenyl)-1-piperazinyl]ethyl]-6-[4-(2-furoyl)-1-piperazinyl]-3(2H)-pyridazinone 6.** This compound was prepared by the same procedure described above, purified by chromatography silica gel using as eluent a stepwise gradient of  $\text{EtOH}$  (0–5%) in  $\text{CH}_2\text{Cl}_2$ ; a dense oil was obtained. Yield: 45%.  $^1\text{H-NMR}$  ( $\text{CDCl}_3$ )  $\delta$ : 2.7–2.9 (6H, m, 4H-piperazinic,  $\text{CH}_2$ ), 3.05–3.15 (4H, m, H-piperazinic), 3.3–3.4 (4H, m, H-piperazinic), 3.85–3.95 (4H, m, H-piperazinic), 4.25 (2H, t,  $J = 6$  Hz,  $\text{CH}_2$ ), 6.5 (1H, m, H-furanic), 6.9–7.4 (7H, m, 4H-aromatic, 2H-pyridazinonic, 1H-furanic), 7.5 (1H, s, H-furanic). The corresponding hydrochloride had mp = 125–128 °C.

**2-[[4-(2-Methoxyphenyl)-1-piperazinyl]propyl]-6-[4-(2-furoyl)-1-piperazinyl]-3(2H)-pyridazinone 7.** This compound was

prepared by alkylation of **11** with 4-(2-methoxyphenyl)-1-(3-chloropropyl)piperazine [12], using the method described above, purified by chromatography silica gel using as eluent a stepwise gradient of  $\text{EtOH}$  (0–10%) in  $\text{CH}_2\text{Cl}_2$ . Yield: 40%.  $^1\text{H-NMR}$  ( $\text{CDCl}_3$ )  $\delta$ : 1.9–2.1 (2H, m,  $\text{CH}_2$ ), 2.5 (2H, t,  $J = 6$  Hz,  $\text{CH}_2$ ), 2.6–2.7 (4H, m, 4H-piperazinic), 3.05–3.15 (4H, m, H-piperazinic), 3.3–3.5 (4H, m, 4H-piperazinic), 3.9 (3H, s,  $\text{OCH}_3$ ), 3.9–4.0 (4H, m, H-piperazinic), 4.15 (2H, t,  $J = 6$  Hz,  $\text{CH}_2$ ), 6.5 (1H, m, H-furanic), 6.8–7.0 (5H, m, 4H-aromatic, 1H-pyridazinonic), 7.05–7.15 (2H, m, 1H-pyridazinonic, 1H-furanic), 7.5 (1H, s, H-furanic). The corresponding hydrochloride had mp = 110–115 °C.

**3-Chloro-6-[4-(2-furoyl)-1-piperazinyl]pyridazine 10**

A mixture of 4.9 g ( $2.7 \times 10^{-2}$  mol) of 1-(2-furoyl)piperazine and 4.47 g ( $3.0 \times 10^{-2}$  mol) of 3,6-dichloropyridazine in 2-butanone containing 10 g of anhydrous potassium carbonate, was heated under reflux with stirring for 17 h. The mixture was filtered hot, and the filtrate was evaporated under reduced pressure. The resulting product was purified by flash chromatography using  $\text{CH}_2\text{Cl}_2/\text{EtOH}$  (95:5). Yield: 56%. Mp = 138–142 °C.  $^1\text{H-NMR}$  ( $\text{CDCl}_3$ )  $\delta$ : 3.7–3.8 (4H, m, H-piperazinic), 3.85–3.95 (4H, m, H-piperazinic), 6.5 (1H, m, H-furanic), 6.9 (1H, d, 1H-pyridazine), 7.1 (1H, m, 1H-furanic), 7.3 (1H, d, H-pyridazine), 7.5 (1H, s, H-furanic).

**6-[4-(2-Furoyl)-1-piperazinyl]-3(2H) pyridazinone 11**

A solution of 2 g ( $5.1 \times 10^{-3}$  mol) of **10** in 30 mL of glacial acetic acid was refluxed for 6 h. The acetic acid was removed under reduced pressure, and the residue dissolved in water and extracted with  $\text{CH}_2\text{Cl}_2$ . The organic phase, dry on  $\text{Na}_2\text{SO}_4$ , was evaporated under reduced pressure. The residue was purified by gel chromatography using as eluent a stepwise gradient of  $\text{EtOH}$  (0–10%) in  $\text{CH}_2\text{Cl}_2$ . Yield: 40%. Mp = 212–215 °C.  $^1\text{H-NMR}$  ( $\text{CDCl}_3$ )  $\delta$ : 3.25–3.4 (4H, m, H-piperazinic), 3.85–3.95 (4H, m, H-piperazinic), 6.5 (1H, m, H-furanic), 6.85 (1H, d,  $J = 9.5$  Hz, 1H-pyridazinonic), 7.1 (1H, m, 1H-furanic), 7.15 (1H, d,  $J = 9.5$  Hz, H-pyridazinonic), 7.5 (1H, s, H-furanic), 10.4 (1H, s, NH-pyridazinonic).

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**References**

- Hieble JP, Bylund DB, Clarke DE et al (1995) *Pharmacol Rev* 47, 267–269
- Gerge P, Borg F, O'Connor S et al (1995) *Eur J Med Chem* 30, 299–303
- Raghupathi RK, Rydelek-Fitzgerald L, Teitler M, Glennon RA (1991) *J Med Chem* 34, 2633–2638
- Price DT, Chari RS, Berkowitz DE, Meyers WS, Schwinn DA (1994) *Mol Pharmacol* 46, 221–226
- Corsano S, Strappaghetti G, Scapicchi R, De Blasi A, Barbarulo D (1995) *2nd Congress Chim Pharm*, Ferrara, August 30–September 3
- Cotecchia S, Schwinn DA, Randall RR, Lefkowitz JR, Caron MG, Kobilka BK (1988) *Proc Acad Natl Sci USA* 85, 7159–7163
- Schwinn DA, Lorenz W, Szklut PJ et al (1990) *J Biol Chem* 265, 8183–8189
- Testa R, Taddei C, Poggiesi E, Destefani C et al (1995) *Pharm Comm* 6, 79–96
- Morrow AL, Creese I (1986) *Mol Pharmacol* 29, 321–330
- Hoyer D, Engel G, Kalkman HO (1985) *Eur J Pharmacol* 118, 13–23
- Lespagnol A, Deprey J (1962) *Bull Soc Chim Fr* 197, 1117–1122
- Bourdais J (1968) *Bull Soc Chim Fr* 8, 3246–3249
- Fratelli M, Marasco O, De Blasi A (1987) *Biochem Biophys Acta* 930, 87–90